
The Use of Nanotechnology to Improve the Neuroprotective Effects of Adenosine in Stroke and Spinal Cord Injury

There remains a significant need to develop successful pharmacological neuroprotective agents for the treatment of...
neurological diseases such as ischemic stroke and spinal cord injury (SCI). Barriers to successful neuroprotective drug development include a high rate of drug metabolism, rapid central nervous system clearance, and difficulty penetrating the blood-brain and blood–spinal cord barriers. By enhancing pharmacokinetic profiles and improving neurovascular access, nanotechnology has been shown both to improve drug efficacy and to promote brain delivery.1 One known neuroprotective agent, adenosine, exerts pleotropic protective functions by decreasing excitatory amino acid release, limiting calcium influx and free radical formation, promoting ATP formation, modulating astrocytic and microglial cells, and minimizing N-methyl-D-aspartate receptors.2 Unfortunately, adenosine is unable to penetrate the blood-brain or blood–spinal cord barrier, has a short half-life, and promotes cardiovascular depression at high doses.

In a recent issue of *Nature Nanotechnology*, Gaudin et al3 generate an intravenous nanoparticle through the bioconjugation of adenosine with squalene (a biocompatible lipid). This squalenoylation technology creates an amphiphilic prodrug that spontaneously forms nanoparticles that are 120 nm in size, allowing prolonged drug interaction with the neurovascular unit while minimizing toxic side effects. In a middle cerebral artery occlusion model, subjects were subjected to 2 hours of occlusion followed by 22 hours of reperfusion. Squalenoyladenosine (SQAd) nanoassemblies (7.5 or 15 mg/kg) were delivered intravenously before ischemia. Infarct volumes were reduced to 31 ± 2 mm³ in the 7.5-mg/kg group and 17 ± 1 mm³ in the 15-mg/kg arm. These volumes were significantly decreased compared with the control arms: 5% dextrose (49 ± 1 mm³), free adenosine (5.5 mg/kg, 55 ± 3 mm³), or unconjugated squalenoyl nanoassemblies (9.45 mg/kg, 44 ± 2 mm³). Figure). The authors went on to show that the timing (2 hours after occlusion or 24 hours after permanent occlusion in the absence of reperfusion) of SQAd delivery did not significantly alter infarct size (23 ± 2 vs 24 ± 4 mm³, respectively). Interestingly, systemic administra-
tion of both adenosine-1 (CCPA) and adenosine-2 (CGS21680) receptor agonists resulted in high mortality owing to adverse cardiac events and did not confer neuroprotection, respectively.3 These results suggest that increased drug bioavailability rather than the timing of drug delivery or direct adenosine receptor activation accounts for the neuroprotective effect observed with the SQAd nanoassemblies.

The pathogenesis of secondary SCI shares many of the same injury mechanisms known to play a role in cerebral ischemia. To assess the effect of SQAd on SCI, behavioral and ultrastructural studies were performed on Sprague-Dawley rats subjected to a T9 transection injury.4 To reproduce an SCI, a 4-g weight was dropped from a height of 12.5 mm after a thoracic laminectomy. SQAd nanoassemblies (32 mg/kg), SQAd nanoassemblies (20.5 mg/kg), free adenosine (11.5 mg/kg), or 5% dextrose was administered 5 minutes after injury. Functional evaluations were performed in a blinded fashion at 24, 48, and 72 hours and 28 days, followed by ultrastructural analysis of a sample of contused tissue 48 hours and 28 days after injury. Animals receiving dextrose or free adeno-
sine were unable to move their hind limbs after injury; on structural assessment, visible contusion of their cord was observed. In the treatment arm, the animals receiving SQAd demonstrated coordinated between their hind limbs and forelimbs during ambulation without a visible contusion on ultrastructural studies. Finally, with the use of the the Baso, Beatti, and Bresnahan grading scale, SQAd animals scored a 14.4, a sign of axonal transduction through a lesion site.

The exact neuroprotective mechanism of these SQAd nanoassemblies remains unclear. The injection of SQAd into the stroke model, ischemic microvessels were found to be patent 6 hours after middle cerebral artery reperfusion, whereas in the control arm, ischemic capillaries were occluded by erythrocytes. Furthermore, SQAd-treated mice lacked swelling of astrocytic end feet around microvessels and endothelial nuclei compared with control mice, an indication that adenosine released from the nanoassemblies penetrates endothelium to interact with the microcirculation and pericyte receptors. Most important, the effective dose of adenosine in the SQAd compound did not cause systemic vasodilation and cardiovascular collapse, a side effect that would have been unavoidable with an unbound nucleoside administered at a neuroprotective dose.

Nanoengineering and squalenoylation techniques have been avidly applied to anticancer and antiretroviral compounds but have not been used less in stroke and SCI. Managing this technology without restricting pharmaceutical design will prove to be a difficult but essential task in advancing the therapeutic use of nanotechnology in neurological diseases.

**REFERENCES**


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**Focused Ultrasound With Microbubbles Increases Temozolomide Delivery in U87 Transfected Mice**

Malignant astrocytomas, particularly glioblastoma multiforme, are among the deadliest of all human cancers, with a median survival of approximately 15 months. Aggressive treatment with maximal resection followed by chemotherapy and radiation has prolonged survival a matter of months.5 The mechanism of resistance of these tumors to traditional treatment strategies is multifactorial, but the inability to effectively deliver therapeutic agents across the blood-brain barrier (BBB) is clearly one of the major causes of treatment failure. As a result, penetration of endothelial tight junctions of the BBB to provide increased drug delivery is an area of intense research.5

In this report, we review a recent study demonstrating increased delivery of temozolomide (TMZ) to implanted glioma tumor lines in mice using focused ultrasound with microbubbles (FUS).6

Initially, the investigators examined the effects of 2- and 5-W delivery of FUS on tumor-free mice. The 2-W delivery of FUS was shown to open the BBB and did not induce damage to normal tissue via hematoxylin and eosin and Evans Blue staining. The 5-W treatments opened the BBB in a wider distribution but caused extravasation of erythrocytes on gross examination and hematoxylin and eosin and Evans Blue staining. For this reason, all FUS treatments in the subsequent experiments were carried out with 2-W strength.

U87 glioma cells were grown in culture and stereotactically implanted into the striatum of 5- to 7-week-old immunocompromised male mice. Thirty-nine mice received TMZ treatment alone, and 43 received TMZ treatment after FUS. Treatment began when the tumor mass was detectable on magnetic resonance imaging, and all mice were sacrificed after treatment. Implanted tumor, contralateral normal brain, and peripheral blood were harvested and analyzed via liquid chromatography/mass spectroscopy to